**Bacteria culture Day**

* Setup traveling tray with:
  + Loops, culture tubes, pipets, pipet bulb, marker, tube rack, media broth
  + gloves, bleach, and EtOH, sample containers
* Clean biohood with bleach then EtOH then clean everything with EtOH before placing it into the hood
* Retrieve microbe:
  + Obtain desired bacteria for testing from glycerol tube.
    - Remove glycerol stock from -80C freezer and transport to biohood frozen (dry ice or liquid N2)
  + Inoculate selected microbe into liquid media (LB/MRS/TSB, dependent upon which one it was isolated on originally).
    - Prepare culture tube with 6mL of broth media
    - With a loop/scrape the glycerol stock tube and place loop into culture tube (repeat as needed).
    - Return stock tube to freezing conditions **QUICKLY**
  + Incubate the sample in the shaking incubator for a 24-hour period at 32℃.
  + Prepare inoculation schedule and sampling schedule

**Inoculation Day**

* After 24-hours, aliquot 200uL from each culture tube into 4 wells of a 96well plate to obtain an absorbance reading.
* Prepare container intended for bacterial inoculation (sampling container)
* Pellet bacteria and resuspend in saline solution
  + Centrifuge culture tubes at 6400 rcf for 10 minutes.
  + Remove supernatant
  + Add \*resuspension liquid into the culture tube
  + Use pipet and mix bacteria pellet of each sample.
* Add the total volume of each resuspended culture tube to the sample container
* Add 6 ml of treatment liquid into the sample container
* Incubate sample container in a 32C incubator, and sample at 2 hours, 24 hours, and 48 hours after initial inoculation.

\*resuspension media could be saline, liquid media, juice, etc. The goal is to resuspend the pellet, so it can be transferred into the treatment container.